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Tachinids as good biocontrol agents of agricultural pests

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ABSTRACT

Tachinids (Diptera: Tachinidae) are good biocontrol agents of agricultural and other crop plants. They are broadly divided into macro egg laying and micro egg laying individuals. Macro egg laying tachinids deposited their eggs directly on the insect pest body while, micro egg laying tachinids laid on the host plants of insect pests. In both cases, after getting the entry into the host body, eggs hatched into maggots and maggots consumed internal tissues of the pest insects by killing them when they came out by breaking the body wall of pest stages like larvae and pupae. Mostly pest larvae have been found attacked by tachinids. The diversity of tachinids and their biocontrol potential is summarized in the paper.

KEY WORDS: Tachinids, diversity, biocontrol potential.

INTRODUCTION

 ${f T}$ he Tachinids are very potential biocontrol agents of several insect pests. They belong to the family Tachinidae of order Diptera which contain about 10,000 species worldwide (Irwin et al., 2003). The tachinids play significant role in regulating herbivore population and stimulating ecological communities, both natural and managed. Up to date, more than 100 species of tachinids have been employed in biological control programmes of agricultural, horticultural and forest crop plants (Sathe, 2014). Tachinids occur throughout the world from nearly all terrestrial environments. Therefore, tachinids are attracting increasing attention of basic ecologists, taxonomists and the workers involved in biological control of insects. Review of literature indicates that the tachinids have been studied by Allen (1995), Belshaw (1993), Crosskey (1976), Feener and Brown (1997), Felland (1990), Monteith (1955), Roth et al. (1982), Shima (1989), Sathe (2012, 2014), Shendage and Sathe (2014 a, b), Sathe et al. (2014 a, b), etc.

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Material and Methods

Biocontrol potential of tachinids from Kolhapur region of India has been studied by making the survey of tachinid parasitoids on insect pests at weekly interval by collecting immature stages of pests such as larvae and pupae. A very large number of pest larvae and pupae were collected from the fields and reared in the laboratory for screening the tachinids from them. Per cent parasitism was decided on the number of hosts parasitized out of hundred individuals. Different crops and pests have been surved for making the records of pests and their parasitoids" by one man one hour search / collection method during the years (2014-2015). The insect pests and the tachinids emerged from the pests have been identified by consulting appropriate literature cited in references.

Results

Result recorded in table-1 and Fig 1-7 indicated that the tachinids attacked on 20 pest insects from Kolhapur region. The maximum 40.00% and minimum 2.00 % parasitism was recorded on *Spodoptera litura* Fab and *Forficula* sp. by macroegg type parasitoids respectively. Both macroegg type and microegg type of tachinids were prevalent in the Kolhapur region of India (Table-1). In general, macroegg type tachinids were dominant in the region wherein they directly laid their eggs into the body of

Table-1: Biocontrol potential of Tachinids on some insect pests

Sr. No	Tachinid Species	Family/Tribe	Eggs on	Pests insect	% Parasitism
1.	Compsilura sp.	Exoristinae (Blondeliini)	Host	Lymantria dispar (<u>L.</u>)	12.00
2.	Eucelatoria sp.	Exoristinae (Blondeliini)	Host	Helicoverpa armigera (Hubner)	27.00
3.	Lixophaga diatraeae (Townsend)	Exoristinae (Blondeliini)	Host	Diatraea saccharalis (Fab.)	17.00
4.	<i>Drino bohemica</i> Mesn.	Exoristinae (Eryciini)	Host	D. saccharalis	5.00
5.	Drino sp.	Exoristinae (Eryciini)	Host	D. saccharalis	10.1
6.	Exorista bombycis (Louis)	Exoristinae (Blondeliini)	Host	Mythimna separata (Walker) Spodoptera litura (Fab.)	14.00 40.00
7.	Exorista mella (Walker)	Exoristinae (Blondeliini)	Host	<i>Grammia</i> sp.	7.00
8.	Leschenaultia sp.	Tachininae (Ernestiini)	Host	Malacosoma sp.	3.00
9.	Linnaemyia comta Fall.	Tachininae (Ernestiini)	Host	Agrotis ipsilon (Hufnagel)	11.00
10.	<i>Triarthria</i> sp.	Tachininae (Loewiini)	Host	Forficula sp.	2.00
11.	Archytas sp.	Tachininae (Tachiniini)	Host	Heliothis sp.	8.00
12.	Carcelia kockiana Townsend	Exoristinae (Eryciini)	Host	Achea janata L.	3.00
13.	Eucelatoria bryani Sabrosky	Exoristinae (Blondeliini)	Host plants	H. armigera	50.00
14.	Eucelatoria sp.	Exoristinae (Blondeliini)	Host plants	Diatraea sp.	48.00
15.	L. diatraeae	Exoristinae (Blondeliini)	Host plants	D. saccharalis	37.00
16	Exorista mella (Walker)	Exoristinae (Blondeliini)	Host plants	<i>Grammia</i> sp.	16.5
17.	Blepharipa sp.	Exoristinae (Goniini)	Host plants	Lymantria dispar L.	13.00
18.	Leschenaultia sp.	Exoristinae (Goniini)	Host plants	Malacosoma sp.	18.00
19.	Patelloa sp.	Exoristinae (Goniini)	Host plants	Malacosoma sp.	29.00
20.	Blepharipa zebina <u>Walker</u>	Exoristinae (Goniini)	Host plants	Antheraea mylitta Drurry	20.00

hosts. While, microegg type tachinids laid their eggs on host plants which entered into the body of pest insects through its food and tachinids developed on the internal tissues of the host body. In tachinids, there were 4 distinct stages of life cycle namely, egg, larvae, pupae and adult. The larval instars ranged from 4 to 5 in different species. The tachinids attacked old larval instars more than the younger instars and relatively larval parasitoids were more dominant than pupal parasitoids in the region.

Discussion

Tachinids are of two categories. In one case they lay eggs directly on the host body and such eggs are larger in size. Another type of tachinids lay their eggs on host plant and their eggs are thus smaller in size. In this category, the eggs entered into host body through mouth or alimentary canal and after hatching the eggs, the larvae started eating internal tissues of the host body. First category tachinids lay their eggs into host body.

The parasitoid larva moults three / four time in host body. The last instar, which is fully matured break the body wall of the host and come out and spin the brownish barrel shaped cocoon. While breaking body wall of host and coming outside the host body, the host dies on the spot. Thus, tachinids help in controlling pest species.

Figure-1. *H. armigera* (polyphagus pests)



Figure-2. Exorista sp. parasitizing H. armigera



Figure-3. S. litura (pest defoliator)



According to Al-Dubai et al. (2012) the sex ratios of flies captured in the flower baited to control traps were subsequently related to flower density, width, depth and plant height in order to test the hypothesis

that flower and plant morphology influenced their attractiveness and / or accessibility to Tachinidae (Fiedler and Landis, 2007, Sivinski et al., 2011). According to Stireman et al. (2006), 909 tachinids were feeding on flowering plants, 28% were Phasiinae.

Figure-4. Exorista sp



Figure-5. Exorista sp



Figure-6. M. separata



There was a substantial evidence that increasing the floral diversity of agroecosystems enhances natural enemy diversity and abundance and ultimately the biological control of pest insects was possible. However, some of the abundantly collected tachinids have contributed to pest suppression. (Al-Dubai et al., 2012) genus Archytas attacked a wide variety of Lepidopteran larvae (Arnaud, 1978) and out of which some use agriculturally more economic important and mass reared for augmentative release (Mannion et al., 1995). The hosts of C. townsendi, led 32% of the capture of Tachinidae. Sathe et al. (2014, 2015) recorded number of biocontrol agents on pest insects found on various agroecosystems.

Finally, it is concluded that the survey and resurveys of tachinids and their biocontrol potential be made frequently and continuously for undating the knowledge of tachinid occurrence in the region and their possible utilization in biological control of insect pests as ecologically approach of pest management. The tachinids are also good pollinating agents for increasing yield of the crops.

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Conflict of Interests

Authors declare that there is no conflict of interests regarding the publication of this paper.

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